

# Quantification of GLP-1R trafficking in primary $\beta$ -cells in response to different ligands

Sarah M. Gray<sup>\*1</sup>, Peter Ravn<sup>2</sup>, Kyle W. Sloop<sup>3</sup>, Jonathan E. Campbell<sup>1,4,5</sup>, David A. D'Alessio<sup>1,5</sup>

<sup>1</sup>Duke Molecular Physiology Institute, Duke University, Durham, NC, USA.

<sup>2</sup>Antibody Discovery and Protein Engineering, BioPharmaceuticals R&D, AstraZeneca, Cambridge, United Kingdom

<sup>3</sup>Diabetes and Complications, Lilly Research Laboratories, Eli Lilly and Company, Indianapolis, IN, USA.

<sup>4</sup>Department of Pharmacology and Cancer Biology, Duke University, Durham, NC, USA

<sup>5</sup>Department of Medicine, Division of Endocrinology, Duke University, Durham, NC, USA

# Disclosures

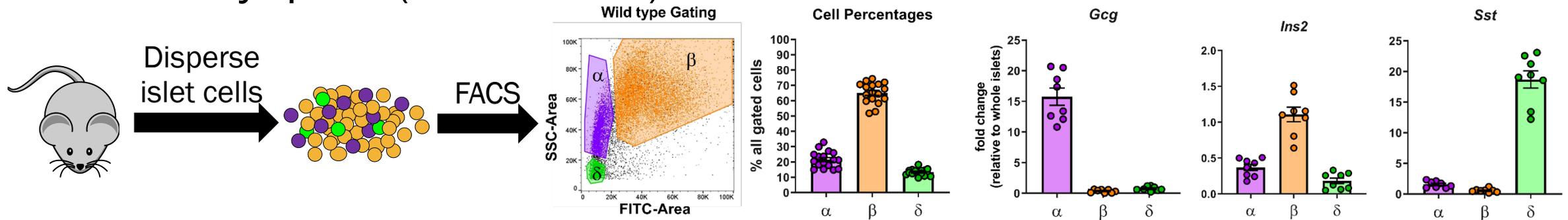
- K.W.S. is an employee of Eli Lilly
- A portion of this work was supported by Eli Lilly and Company through the Lilly Research Award Program (LRAP)

# Introduction

- The glucagon-like peptide 1 receptor (GLP-1R) is a class B G-protein coupled receptor expressed in islet  $\beta$ -cells. Activation of the GLP-1R is insulinotropic.
- Multiple endogenous and synthetic ligands exist for the GLP-1R. Emerging evidence suggests that each ligand has a unique interaction with GLP-1R, causing distinct responses of receptor physiology.
- In these studies, we test the hypothesis that specific GLP-1R ligands differentially affect GLP-1R trafficking and internalization in primary  $\beta$ -cells.

# Methodology

- Enriched populations of  $\beta$ -cells were generated by flow cytometry/FACS and validated by qPCR (see 2113-P)



- Glp1r0017, an antibody for GLP-1R (Biggs et al., *Diabetologia* 2018), was conjugated to APC (GLP1R-APC) and validated for use in flow cytometry applications (see 2113-P)
- Dispersed islet cells were treated with GLP-1R ligands, incubated with GLP1R-APC, and  $\beta$ -cell staining was analyzed on a flow cytometer (Attune NxT Analyzer) or imaged by ImageStream X

# Ligand exposure decreases GLP1R-APC Staining in $\beta$ -cells and suggests internalization

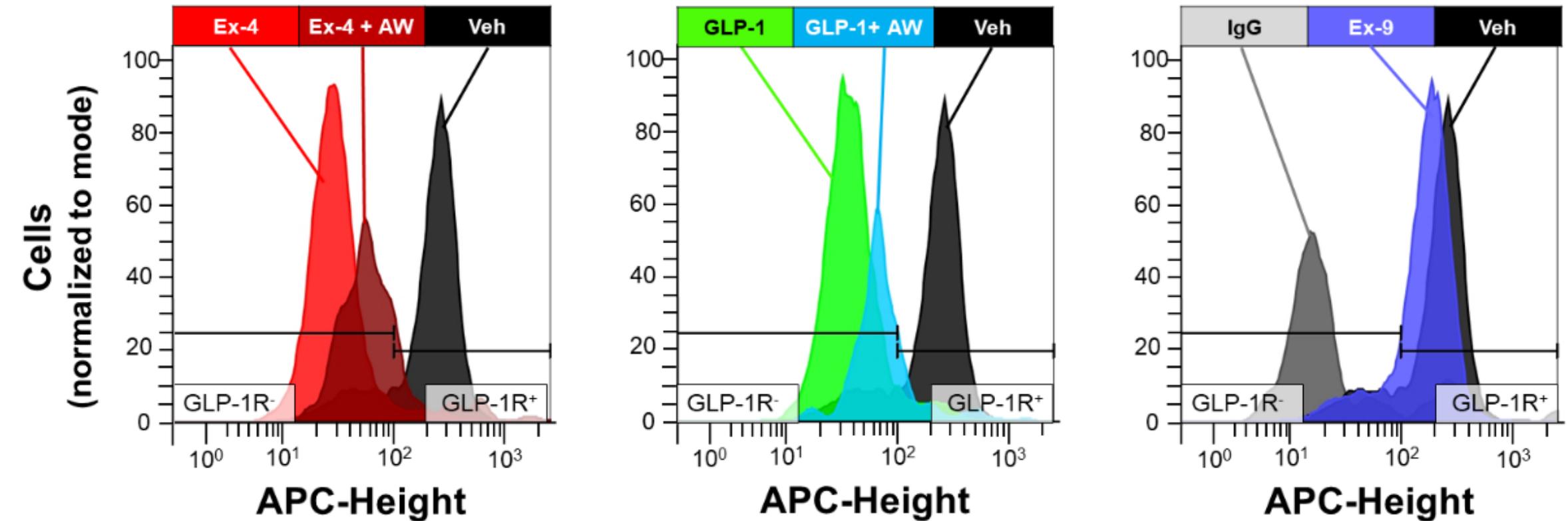
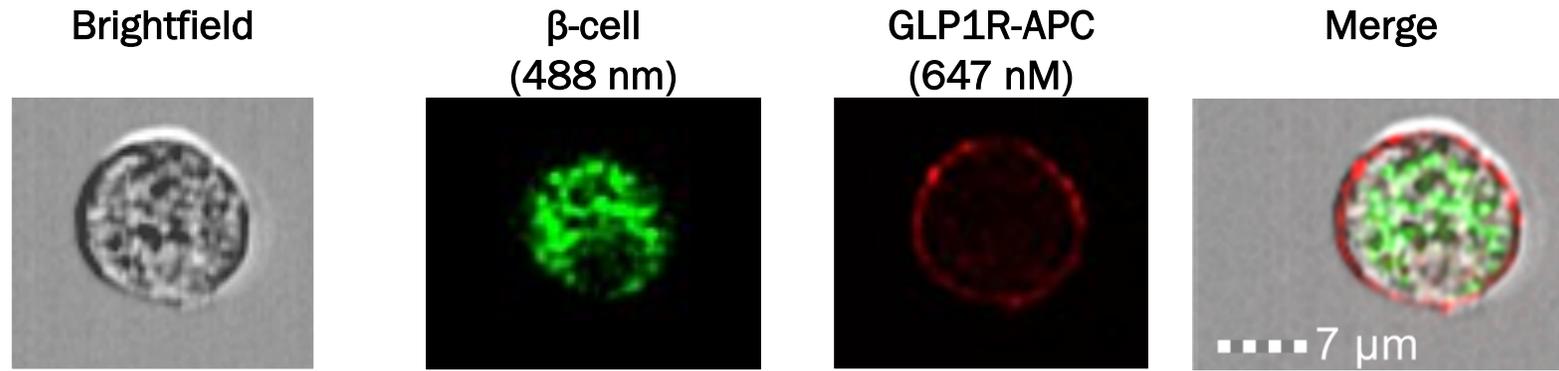


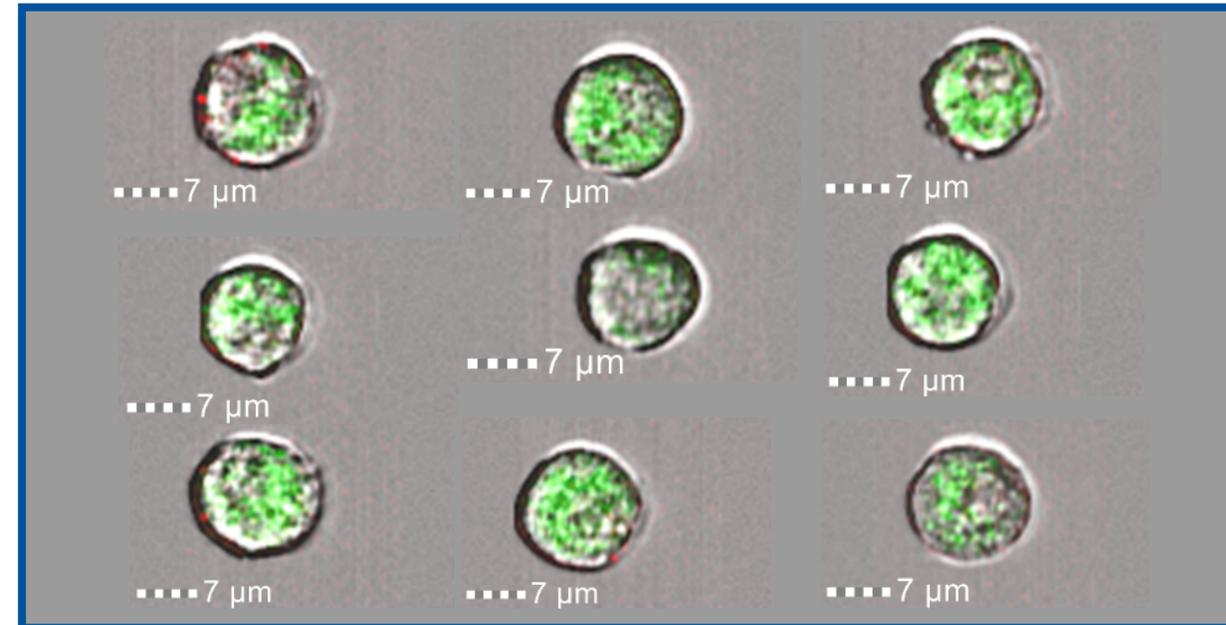
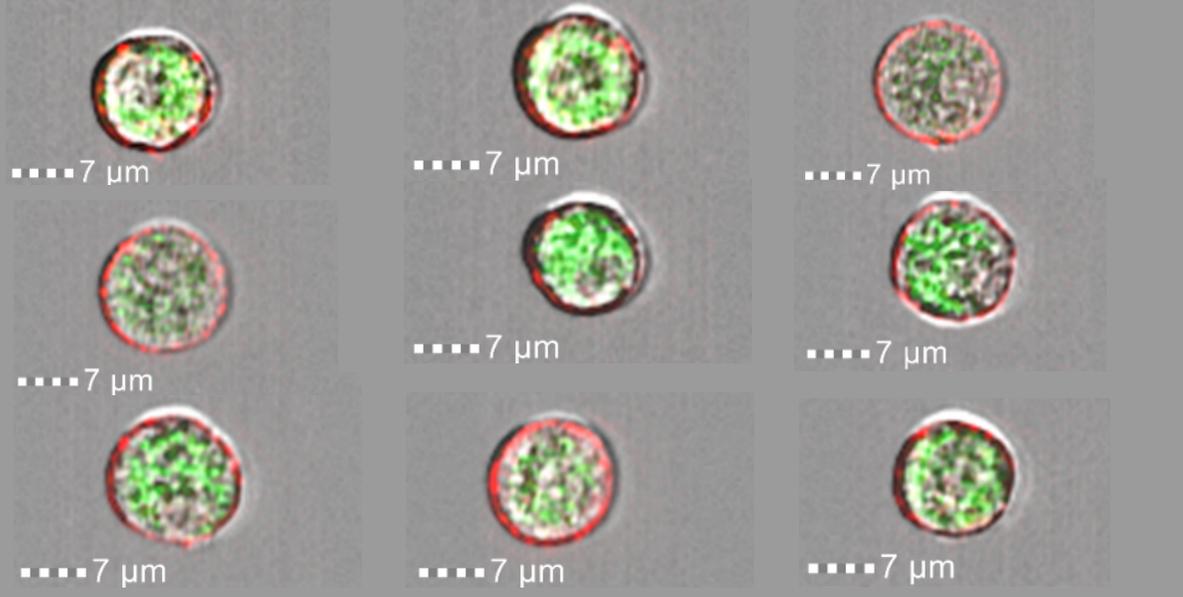
Figure 1. Following ligand treatment, GLP-1R is internalized and there is a left-shift in GLP1R-APC staining. Acid wash does not significantly alter this, suggesting ligands do not interfere with GLP1R-APC Staining. Histogram of  $\beta$ -cells stained with GLP1R-APC following A) exendin-4 alone (Ex-4) or followed by acid wash (Ex-4 + AW), or vehicle (Veh), B) GLP-1 alone (GLP-1) or GLP-1 followed by acid wash, or vehicle, C) exendin 9-39 (Ex-9) or vehicle, with IgG-APC (IgG) shown. All ligands were given at 100 nM for 30 min at 37°C. Acid wash conditions followed ligand treatment (2x wash with 0.5M NaCl, 0.2 M acetic acid, pH 2.5). Following treatment, reactions were stopped on ice and incubated with GLP1R-APC or IgG-APC.

# Ligand exposure reduces GLP1R-APC Staining



Vehicle

Ex-4 (100 nM, 30 min, 37°C)



**Figure 2.** Imagestream allows for combined microscopy and flow cytometry and demonstrates decreased GLP1R-APC staining in  $\beta$ -cells following Ex-4 treatment. **Top:** Representative images from brightfield, 488 nm, and 647 nm channels with merged image for a single  $\beta$ -cell. **Bottom:** GLP-1R staining in  $\beta$ -cells treated with vehicle or exendin-4 (Ex-4). Autofluorescence (488, green) was used to select  $\beta$ -cells and APC (red) was used to visualize GLP-1R.

# Exposure to ligands causes a dose-dependent reduction of GLP-1R on $\beta$ -cell plasma membrane

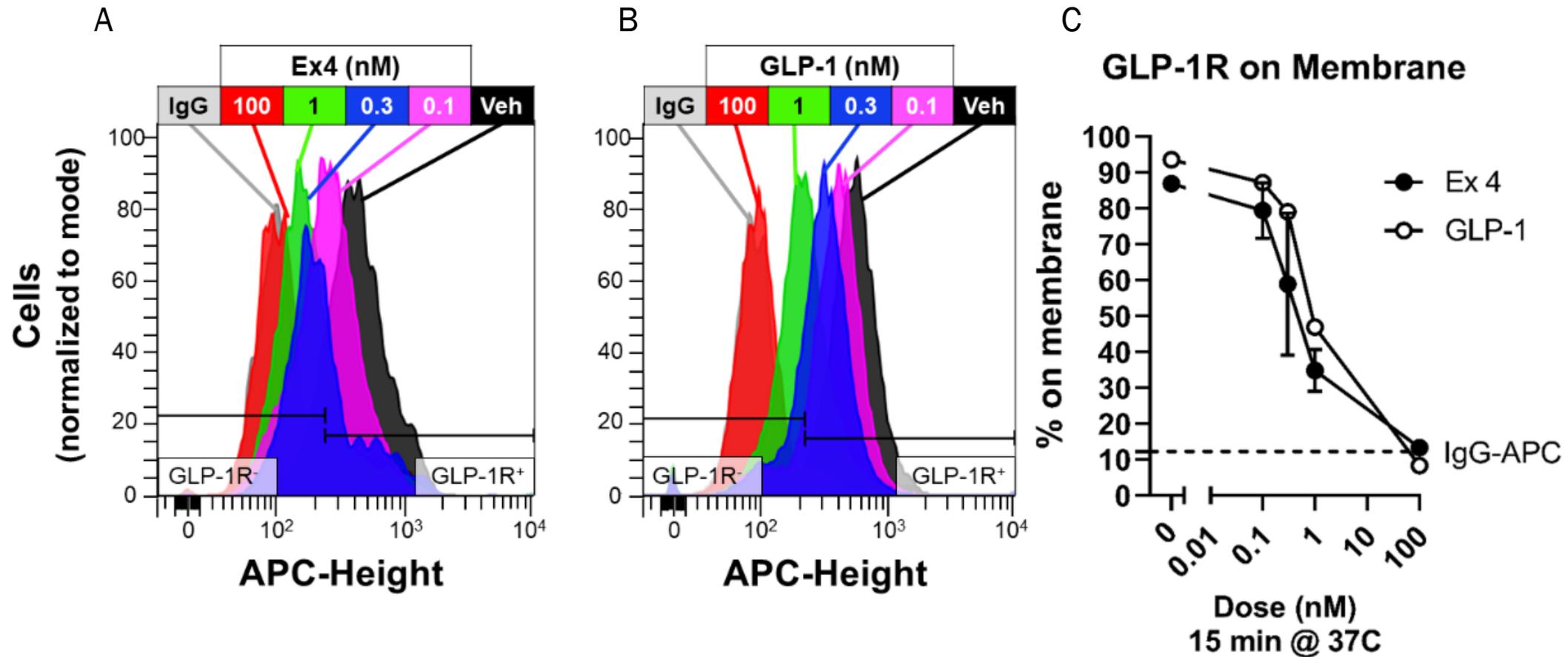


Figure 3. Increasing doses of GLP-1R ligand caused a shift in GLP1R-APC staining from normal (Veh) to absent (IgG), suggesting progressive internalization in response to ligand exposure. Histograms of GLP1R-APC staining following increasing doses of A) exendin-4 (Ex4) and B) GLP-1 with C) quantification. Dispersed cells were treated for 15 min at 37°C with ligand or vehicle prior to GLP1R-APC staining. N=1-2 replicates per ligand.

# Exendin-4 promotes GLP-1R internalization in proportion to time of exposure

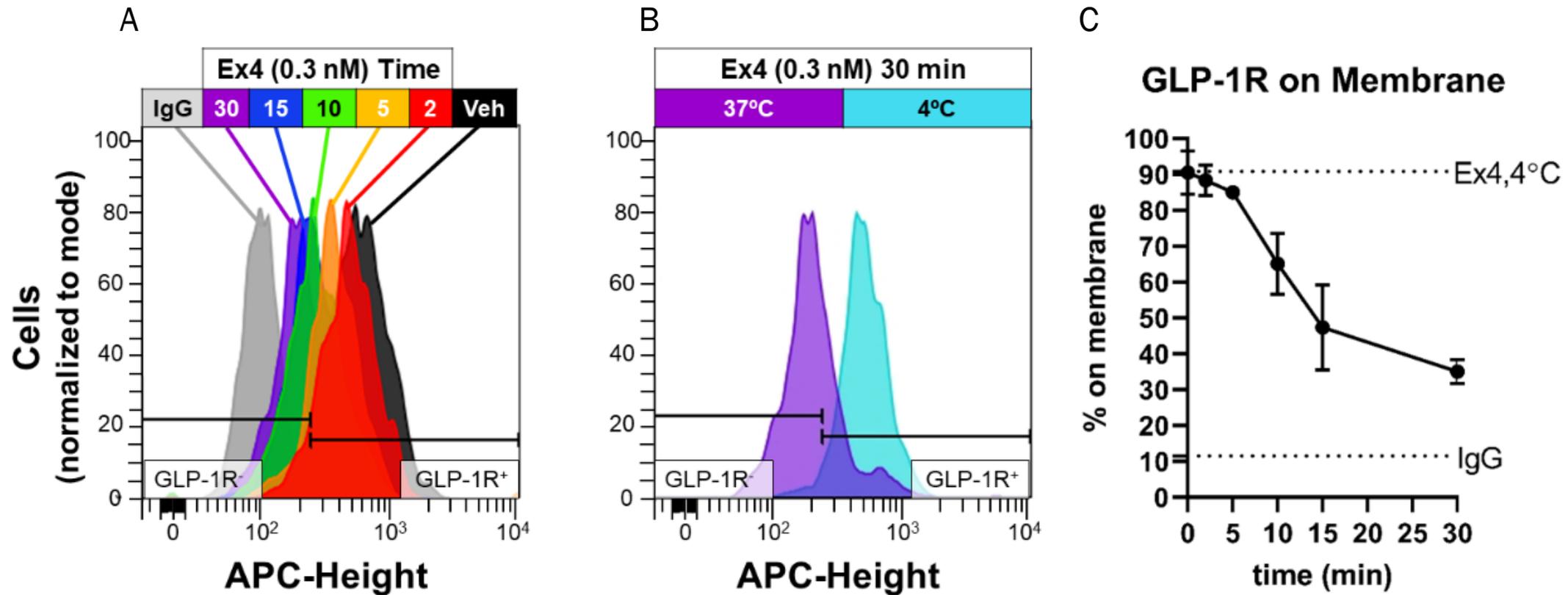


Figure 4. Increased time of exposure to Ex-4 shifted GLP1R-APC staining from normal (veh) toward absent (IgG) and this response was muted at 4°C. Histograms of GLP1R-APC staining following A) Time course of exendin-4 (Ex4, 0.3 nM) and B) 30 min at 37°C or 4°C with C) quantification. Dispersed cells were treated for with 0.3 nM Ex4 at 37°C or 4°C with Ex4 or vehicle prior to GLP1R-APC staining. N=2.

# GLP-1R recycles slowly after maximal Exendin-4 dose

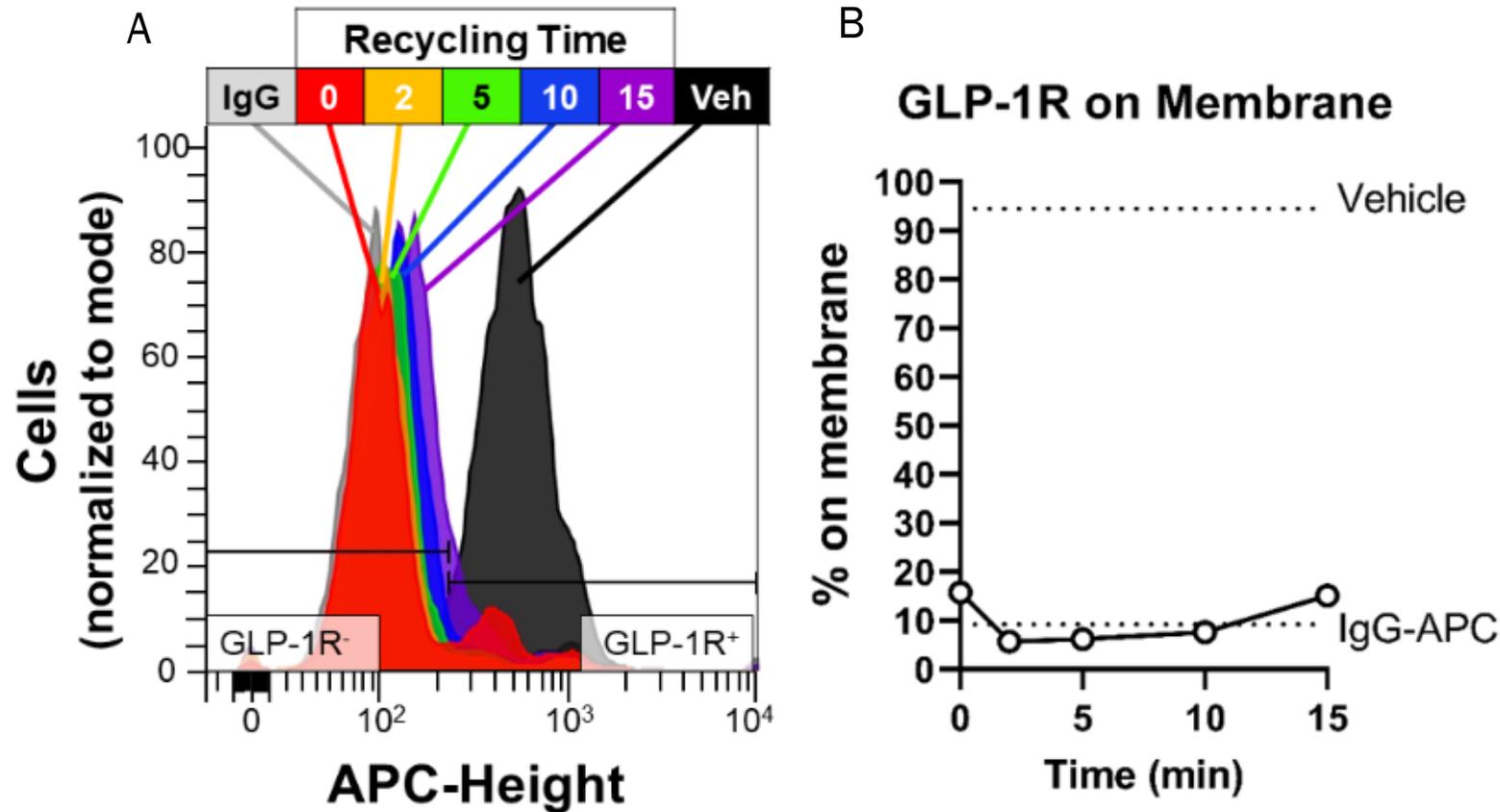


Figure 5. Following exposure to Ex-4, there was a modest right-shift of GLP1R-APC staining for 15 minutes after exposure, consistent with slow recycling of GLP-1R after maximal ligand exposure and internalization. Dispersed islet cells were treated with Ex4 at 100 nM for 30 min at 37°C before ligand was washed out and cells were incubated in buffer at 37°C for specified amount of time to allow GLP-1R to recycle to plasma membrane. A) Time course of GLP-1R recycling after Ex4 and B) quantification. Dispersed cells were treated for with 0.3 nM Ex4 at 37°C or 4°C with Ex4 or vehicle prior to GLP1R-APC staining. N=1.

# Conclusions

- Conjugation of Glp1r0017 to a fluorophore (GLP1R-APC) allows for imaging of the GLP-1R during flow cytometry applications in live cells
- GLP1R-APC binding allows for inferences about GLP-1R internalization
- GLP1R-APC binding is not prevented by ligand treatment
- Following maximal Ex4 treatment, GLP-1R on plasma membrane is undetectable and slow to recycle back to the plasma membrane
- Differences in dose- and time-dependent movement of GLP-1R in response to GLP-1R ligands requires further investigation